This article was downloaded by:

On: 26 January 2011

Access details: Access Details: Free Access

Publisher Taylor & Francis

Informa Ltd Registered in England and Wales Registered Number: 1072954 Registered office: Mortimer House, 37-

41 Mortimer Street, London W1T 3JH, UK



Nucleosides, Nucleotides and Nucleic Acids

Publication details, including instructions for authors and subscription information: http://www.informaworld.com/smpp/title~content=t713597286

Stereospecific Synthesis and Anti-HIV Activity of (Z)2'- and (E)3'-Deoxy-2'(3')-C-(chloromethylene) Pyrimidine Nucleosides

Elena N. Kalinichenko^a; Elena B. Rubinova^a; Evgueny V. Borisov^a; Jan Balzarini^b; Erik De Clercq^b; Igor A. Mikhailopulo^a

^a Institute of Bioorganic Chemistry, Byelorussian Academy of Sciences, Minsk, Byelorussia ^b Katholieke Universiteit Leuven, Rega Institute for Medical Research, Leuven, Belgium

To cite this Article Kalinichenko, Elena N. , Rubinova, Elena B. , Borisov, Evgueny V. , Balzarini, Jan , De Clercq, Erik and Mikhailopulo, Igor A.(1995) 'Stereospecific Synthesis and Anti-HIV Activity of (Z)2'- and (E)3'-Deoxy-2'(3')-C-(chloromethylene) Pyrimidine Nucleosides', Nucleosides, Nucleotides and Nucleic Acids, 14:3,533-536

To link to this Article: DOI: 10.1080/15257779508012420 URL: http://dx.doi.org/10.1080/15257779508012420

PLEASE SCROLL DOWN FOR ARTICLE

Full terms and conditions of use: http://www.informaworld.com/terms-and-conditions-of-access.pdf

This article may be used for research, teaching and private study purposes. Any substantial or systematic reproduction, re-distribution, re-selling, loan or sub-licensing, systematic supply or distribution in any form to anyone is expressly forbidden.

The publisher does not give any warranty express or implied or make any representation that the contents will be complete or accurate or up to date. The accuracy of any instructions, formulae and drug doses should be independently verified with primary sources. The publisher shall not be liable for any loss, actions, claims, proceedings, demand or costs or damages whatsoever or howsoever caused arising directly or indirectly in connection with or arising out of the use of this material.

STEREOSPECIFIC SYNTHESIS AND ANTI-HIV ACTIVITY OF (Z)2'- AND (E)3'-DEOXY-2'(3')-C-(CHLOROMETHYLENE) PYRIMIDINE NUCLEOSIDES

Elena N. Kalinichenko¹, Elena B. Rubinova¹, Evgueny V. Borisov¹, Jan Balzarini², Erik De Clercq², and Igor A. Mikhailopulo¹*

¹Institute of Bioorganic Chemistry, Byelorussian Academy of Sciences, 220141 Minsk, Byelorussia and ²Katholieke Universiteit Leuven, Rega Institute for Medical Research, Minderbroedersstraat 10, B-3000, Leuven, Belgium

Abstract. Reaction of 1-[2,5(and 3,5)-di-O-trityl-β-D-erythro-pentofuran-3(and 2)-ulosyl]uracil derivatives 5 and 6 with (chloromethyl)triphenylphosphorane resulted in the stereoselective formation of (E)-3'- and (Z)-2'-chloromethylene derivatives 7 (69%) and 8 (53%), respectively, deprotection of which gave 9 and 10. Transformation of the uracil nucleoside 7 into cytosine one followed by deprotection yielded 12. The latter was converted into the arabinoside 14. The fully deprotected chloromethylene nucleosides were tested for their activity against HIV-1 and HIV-2.

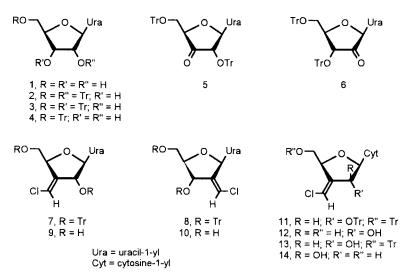
In the last few years, the 2'- and 3'-methylenenucleoside analogues have received considerable attention as valuable synthetic intermediates $(e.g., \ ^1)$ and as potential chemotherapeutic agents $(e.g., \ ^2)$ as well. The present report deals with the preparation and anti-HIV testing of (E)-3'-deoxy-3'-C-(chloromethylene) derivatives of uridine (9), cytidine (12), and ara-C (14) as well as (Z)-2'-deoxy-2'-C-(chloromethylene)uridine (10).

Tritylation of uridine (TrCl, DMAP, pyridine) followed by silica gel column chromatography afforded 2,5- and 3,5-di-O-substituted derivatives 2 (41%) and 3 (29%) as well as 5'-O-trityluridine (4) (18%). Compounds 2 and 3 were oxidized with CrO₃/pyridine/Ac₂O complex³ and the respective ketones 5 and 6 thus obtained condensed with (chloromethyl)triphenylphosphorane generated from (chloromethyl)triphenylphosphonium chloride and BuLi in THF at -30 °C to give, after deblocking and chromatography, 9 and 10 in good combined yields. Only one geometric isomer of both 9(E) and 10(Z) was observed by ¹H NMR spectroscopy.

Assignment of all protons of fully deprotected nucleosides 9 and 10 was achieved by a combination of two-dimensional ¹H, ¹H-COSY and NOESY experiments (data not shown). The stereochemistry of vinyl chlorides 9 and 10 was established by NOE difference spectroscopy. Comparison of NOE magnitudes for the vinylic proton H6' upon irradiation of H2' and H4' (3.8 and 1.2%, respectively) unequivocally supports the E configuration of 9. Moreover, saturation of H2' results in NOE at H6 of 15.4% pointing to the

predominant *anti* conformation of the base at the glycosidic bond. The following data are most informative in the case of 10: $f_{H6}(H3')$ and $f_{H3}(H6')$ are found to be 4.6 and 1.5%, respectively; no NOE at H6' was observed upon irradiation of H1' and *vice versa*. The results in NOE's at H6 of 6.9, 7.7 and 1.1% were obtained upon saturation of H1', H3' and H6', respectively. The latter data point to the predominant high-*anti* conformation of the base at the glycosidic bond. The population of the high-*anti* conformation in 10 suggests an interaction of π electrons of C5-C6 double bond and unsaturated function of the sugar. Indeed, the shielding of H6 resonance (0.43 ppm) in going from 9 to 10 reflects this interaction. Moreover, 10 displays negative Cotton effect at 259 nm, while uridine shows positive band in the B₂₀ region.

2',5'-Di-O-tritylated derivative of 7 was converted into the cytosine vinyl chloride 12 by successive treatment with POCl₃/1,2,4-triazole, conc. ammonia in 1,4-dioxane⁴, and 80% aq. AcOH (55%; combined). Arabinoside 14 was obtained by selective 5'-O-tritylation of 12 to afford 13, followed by reaction with DAST, O²,2'-anhydro ring cleavage under basic conditions, and detritylation (47%; combined).



None of the fully deprotected chloromethylene nucleosides showed activity against HIV-1 and HIV-2 in human T-lymphocyte (CEM/0) cells.

Experimental Section

Thin layer chromatography (TLC) was carried out on the F 1500 LS 254 silica gel plates (Schleicher & Schull, Germany). As solvent systems were used: ethyl acetate-hexane, 1:2 (A); ethyl acetate-hexane, 7:5 (B); chloroform-methanol, 9:1 (C); chloroform-methanol, 4:1 (D), n-butanol-acetic acid-water, 5:3:2 (E). Column chromatography was performed on silica gel L $40/100~\mu$ and $100/400~\mu$ (Chemapol, Czechoslovakia). The solutions of compounds in organic solvents were dried with anhydrous sodium sulfate for 4 h. The reactions were performed at room temperature, unless stated otherwise. All new compounds synthesized gave satisfactory FAB MS and elemental analysis.

1-[2,5(and 3,5)-di-O-trityl-β-D-erythro-pentofuran-3(and 2)-ulosyl]uracils (5 and 6) were prepared³ from 2 and 3 in yields of 89 and 85%, respectively; mp's and ¹H NMR spectra for both compounds were in accord with previously published data⁵. 1-[2,5(and 3,5)-di-O-trityl-3(and 2)-deoxy-3(and 2)-chloromethylene-β-D-erythro-pentofurano-syl]-[3'E(and 2'Z]uracils (7 and 8). A solution of butyllithium (1.5 M in hexane, 0.91 mL, 1.375 mmol) was added dropwise over 10 min to a suspension of (chloromethyl)triphenylphosphonium chloride (0.56 g, 1.65 mmol) in anh. THF (2 mL) at -30 °C under argon. The resulting clear, bright yellow mixture was stirred at 0 °C for 0.5 h, cooled to -30 °C and then a solution of 5 (0.2 g, 0.275 mmol) in anh. THF (3 mL) was added slowly. The mixture was allowed to warm to room temperature and after 2 h saturated NH₄Cl-H₂O was added. The aqueous phase was extracted with ethyl acetate (3x30 mL) and organic extracts were combined, dried and evaporated. The residue was chromatographed, using a linear ethyl acetate gradient (10→60%,v/v, 2 x 500 mL) in hexane to yield 0.15 g (69%) of 7 as a foam which was crystallized from ethyl acetate with hexane added until slight turbidity. Mp >300 °C (decomp.); R_f 0.35 (A); UV (MeOH) λ_{max} nm (ε): 261 (17,900), λ_{min} nm (ε): 245 (9,900).

In a similar way, starting from 6 (0.15 g, 0.206 mmol), 85 mg (53%) of 8 was prepared as an amorphous powder. Mp >300 °C (decomp.); R_f 0.28 (A); UV (MeOH) λ_{max} nm (ϵ): 260 (13,800), λ_{min} nm (ϵ): 245 (9,400).

[3'E(and 2'Z)]-1-(3(and 2)-deoxy-3(and 2)-chloromethylene-β-D-erythro-pentofuranosyl)uracils (9 and 10). A suspension of 7 (0.26 g, 3.42 mmol) in AcOH (80%, 20 mL) was heated at 80 °C for 6 h and evaporated. The residue was coevaporated with toluene and purified by column chromatography, using a linear methanol gradient (0 \rightarrow 10%, v/v, 2 x 250 mL) in chloroform. The product containing fractions were collected, evaporated, dissolved in chloroform (0.1 mL) and precipitated in hexane (100 mL) to give 75 mg (80%) of 9 as an amorphous powder. Mp 180-182 °C; R_f 0.18 (C); UV (MeOH) λ_{max} nm (ε): 260 (11,200), λ_{min} nm (ε): 230 (2,100); CD (MeOH), λ nm ([Θ]·10⁻³): 268 (+5.48), 240 sh (-10.3), 214 (-27.4), λ nm, [Θ] = 0: 282, 257. ¹H NMR (DMSO-d₆): 7.91 (d, 1H, J_{5,6} = 8.0; H-6), 6.24 (t, 1H, J_{2',6'} = J_{4',6'} = 2.25; H-6'), 6.13 (d, 1H, J = 6.0; 2'-OH), 5.70 (d, 2H; J_{1',2'} = 8.0; H-1' and H-5), 5.24 (br.s, 1H; 5'-OH), 4.73 (br.t, 1H, J_{4',5'} = 2.25, J_{4'5''} = ~1.0; H-4'), 4.57 (m, 1H, J_{2',4'} = 2.25, H-2'), 3.74 (dd, 1H, J_{5',5''} = 11.0; H-5').

In a similar way, starting from **8** (0.23 g, 0.303 mmol), 46 mg (55%) of **10** was prepared. Mp 122-124 $^{\circ}$ C (from MeOH); R_f 0.13 (A); UV (MeOH) λ_{max} nm (ϵ): 259 (11,300), λ_{min} nm (ϵ): 230 (1,900); CD (MeOH), λ nm ($\{\Theta\}$ -10-3): 259 (-15.5), λ nm, $\{\Theta\}$ = 0: 285. 1 H NMR (DMSO-d₆): 7.48 (d, 1H, J_{5,6} = 8.0; H-6), 6.52 (t, 1H; J_{1',3'} = 2.25; H-1'), 6.49 (t, 1H, J_{1',6'} = J_{3',6'} = 2.25; H-6'), 5.64 (d, 1H, H-5), 5.90 (d, 1H, J = 6.5; 3'-OH), 4.52 (t, 1H; J = 5.5; 5'-OH), 4.64 (br.t, 1H, H-3'), 3.75-3-52 (m, 3H; H-4', H-5' and H5").

(3'E)-1-(3-deoxy-3-chloromethylene- β -D-*erythro*-pentofuranosyl)cytosine (12). Compound 7 was converted into 11 essentially as described⁴ in 79% combined yield. Mp 260-262 °C (from chloroform-MeOH, 1:1); R_f 0.32 (C).

Compound 11 was detritylated as above to give 12 as acetate in 65%. Mp 118-120 $^{\circ}$ C; R_f 0.31 (D); UV (MeOH) λ_{max} nm (ϵ): 271 (9,500), λ_{min} nm (ϵ): 256 (8,400).

The solution of 12 (acetate) (26 mg, 0.078 mmol) in a mixture of methanol- H_2O (1:2) was applied to a column of Dowex 1x2 (OH⁻-form), which was eluted with methanol- H_2O (1:1, 100 mL). Appropriate fractions were collected, evaporated, dissolved in methanol (0.1 mL) and precipitated in ether (100 mL) to give 23 mg (96%) of 12 as a free base. Mp 114-117 °C; R_f 0.31 (D); UV (MeOH) λ_{max} nm (ϵ): 271 (9,700), λ_{min} nm (ϵ): 256 (8,300); CD (MeOH), λ nm ([Θ]·10⁻³): 270 (+6.0), 220 (-11.5), λ nm, [Θ] = 0: 315, 242. ¹H NMR (DMSO-d₆): 7.83 (d, 1H, J_{5,6} = 7.5; H-6), 7.36 (br.d, 2H, NH₂), 6.26 (t, 1H, J_{2',6'} = J_{4',6'} = 2.5; H-6'), 6.02 (d, 1H, J = 6.5; 2'-OH), 5.81 (d, 1H, H-5), 5.72 (d, 1H, J_{1',2'} = 7.8; H-1'), 5.19 (t, 1H, J = 5.7; 5'-OH), 4.71 (t, 1H, J_{4',2'} = 2.5; H-4'), 4.63 (m, 1H; H-2'), 3.67 (m, 2H; H-5' and H-5").

(3'E)-1-(5-O-Trityl-3-deoxy-3-chloromethylene- β -D-erythro-pentofuranosyl)cytosine (13). A mixture of 12 (0.06 g, 0.22 mmol), triphenylmethyl chloride (0.122 g, 0.44 mmol) and DMAP (1 mg, 0.002 mmol) in anh. pyridine (1 mL) was kept for 2 days and then poured into a mixture of ice and water (100 mL). The resulting precipitate was filtered off, dried and chromatographed with a linear methanol gradient (0 \rightarrow 15%, v/v, 0.5 L) in chloroform to yield 90 mg (78%) of 13 as an amorphous powder. Mp 130-132 °C; R_f 0.43 (C); UV (MeOH) λ_{max} nm (ϵ): 271 (11,700), λ_{min} nm (ϵ): 255 (10,600).

(3'E)-1-(3-deoxy-3-chloromethylene-β-D-arabinopentofuranosyl)cytosine (14). Compound 13 (120 mg, 0.226 mmol) was dissolved in dioxane (1 mL) and (diethylamino)sulfur trifluoride (67 μL, 0.451 mmol) was added under argon. After stirring for 1 h, the reaction mixture was diluted with H₂O (1 mL) and neutralized (pH 7) with BaCO₃. The powder of BaCO₃ was filtered off, washed 3 times with di-

oxane and the filtrate was evaporated to dryness. The precipitate was dissolved in a mixture of chloroform-methanol (0.1 mL) and precipitated in ether (100 mL) to give 100 mg (92 %) of 1-(5-O-trityl-3-deoxy-3-chloromethylene-O²,2-anhydro)-(3'E)-cytosine as an amorphous powder, which was used in the next step without purification; mp >236 °C; R_f 0.58 (E); UV (MeOH) λ_{max} nm (ϵ): 263 (6,700), λ_{min} nm (e): 248 (5,000). The crude product was suspended in AcOH (80%, 10 mL) and the mixture was stirred for 2 h. The solvent was evaporated, and the residue was dissolved in ethanol (4 mL) and treated with 1N NaOH solution (2 mL) for 20 h. The reaction mixture was neutralized with AcOH in ethanol (2.3 mL, 1:1, v/v) and evaporated. The residue was purified with column chromatography, using a linear methanol gradient (5 \rightarrow 20%, v/v, 2 x 300 mL) in chloroform to yield 34 mg (63%) of 14 as acetate. Mp 147-149 °C; R_f 0.37 (D). The solution of 14 (acetate) (34 mg, 0.102 mmol) in a mixture of methanol/H₂O (1:2) was applied to a column of Dowex 1x2 (OH) (10 mL), which was eluted with H₂O (100 mL). The product containing fractions were collected, evaporated, dissolved in methanol (0.1 mL) and precipitated in ether (100 mL) to give 28 mg (91%) of 14 as a free base. Mp 129-131 $^{\circ}$ C; R_f 0.37 (D); UV (MeOH) λ max nm (ε): 273 (13,300), $λ_{min}$ nm (ε): 253 (10,000); CD (MeOH), λ nm ([Θ]·10⁻³): 270 (+20.2), 216 (-¹¹¹¹¹
68.4), λ nm, [Θ] = 0: 300, 235. ¹H NMR (DMSO-d₆): 7.72 (d, 1H, J_{5,6} = 7.5; H-6), 7.10 (br.d, 2H, NH₂), 6.79 (d, 1H, J_{4',6'} = 2.2; H-6'), 5.69 (d, 1H, H-5), 5.82 (d, 1H, J_{1',2'} = 3.0; H-1'), 5.51 (d, 1H, J = 5.8; 2'-1). OH), 5.40 (t, 1H, J = 5.5; 5'-OH), 4.71 (m, 1H, 1H-4'), 4.44 (dd, 1H; 1H-2'), 3.71 (m, 2H; 1H-5'').

Acknowledgement. I.A.M. is deeply grateful to the Alexander von Humboldt-Stiftung (Bonn - Bad-Godesberg, Germany) for partial financial support.

REFERENCES

- Samano, V.; Robins, M.J. Can. J. Chem. 1993, 71, 186-191.; Czernecki, S.; Ezzitouni, A. Tetrahedron Lett. 1993, 34, 315-318.; Svendsen, M.L.; Wengel, J.; Dahl, O.; Kirpekar, F.; Roepstorff, P. Tetrahedron 1993, 49, 11341-11352.; Ioannidis, P.; Classon, B.; Samuelsson, B.; Kvarnström, I. Nucleosides & Nucleotides 1993, 12, 449-462.; Hassan, A.E.A.; Shuto, S.; Matsuda, A. Nucleosides & Nucleotides 1994, 13, 197-211.
- Machida, H.; Sakata, S.; Ashida, N.; Takenuki, K.; Matsuda, A. Antiviral Chem. Chemother. 1993, 4, 11-17.; Matthews, D.P.; Persichetti, R.A.; Sabol, J.S.; Stewart, K.T.; McCarthy, J.R. Nucleosides & Nucleotides 1993, 12, 115-123.; Robins, M.J.; Samano, V.; Zhang, W.; Balzarini, J.; De Clercq, E.; Borchardt, R.T.; Lee, Y.; Yuan, C.-S. J. Med. Chem. 1992, 35, 2283-2293.
- 3. Hansske, F.; Madej, D.; Robins, M.J. Tetrahedron 1984, 40, 125-135.
- 4. Divakar, R. J.; Reese C. B. J. Chem. Soc., Perkin Trans. I. 1982, No. 5, 1171-1176.
- 5. Cook, A.F.; Moffatt, J.G. J. Am. Chem. Soc. 1967, 89, 2697-2705.